

## FLSC Standard Operating Procedure for the Preparation of Immunocompromised Mouse Caging

### Supplies:

Innovive® cages	Innovive® cage bonnet filter top
Sterile corncob bedding 1/4"/Alpha-dri 1:1 mix	Innovive® feeder
Sterile Enviro-dri®	Innovive® prefilled water bottles
Sterile stainless steel cup	Tyvek® autoclave bag
Plastic bags	Rubber bands
Specialty bedding per PI request	Autoclave tape

### Innovive® Cage Preparation

1. All persons handling sterile supplies are required to wear sterile gowns, bouffant caps, particulate filter masks, and sterile gloves and maintain aseptic technique.
2. Place sterilized supplies into the prepared hood and spray the outer surfaces with 70% ethyl alcohol prior to sterile gowning and gloving.
3. Once alcohol has evaporated, the plastic covering Innovive® caging components is opened; the aluminum foil is removed from the autoclaved bedding and environmental enrichment products.
4. Using the sterile cup, place one and a quarter cups of bedding into the cage along with a generous handful of Enviro-dri®.
5. Bed all cages and restack into the sterile plastic bag. Any cage that touches the bench in the hood cannot be stacked into another cage. That cage must be placed into a separate plastic bag.
6. Cages are removed as needed for changing maintaining the sterility of the unused caging.
7. Feeders and cage bonnet tops are opened as needed. Unused feeders are placed into the sterile plastic sleeve with unused cage tops.
8. All opened plastic sleeves containing sterile caging components are twisted closed and secured with a rubber band prior to removal from the sterile hood.
9. Innovive® water bottles are sprayed with alcohol before opening the plastic wrap. The lid opening is covered with an adhesive sticker which must be peeled off for water access.
10. Place unused water bottles into a sterile cage or sterile plastic sleeve for storage.

### Vent Rack or Shoebox Cage Preparation

1. Cages are bedded and environmental enrichment inserted into the cage.
2. Cage tops and bonnets are fitted on the cages. Be sure the filter on the cage bonnet is intact and properly seated.
3. Investigators requesting sterile water in glass bottles will require tall bonnets on the cages.
4. The cage is placed in a Tyvek® bag and the bag closed by pulling the drawstrings tight.
5. Tie the drawstrings in a bow to secure the Tyvek® bag.
6. Place a strip of autoclave tape over the drawstrings containing the date the cage is autoclaved.

### Autoclaving

1. Place the bagged cages onto the autoclave carriage allowing about a finger width between cages. If a second row is needed, arrange the cages so that they are not directly stacked on top of the bottom row.
2. Set the autoclave at 121°C for an exposure time of 15 minutes with an exhaust time of 5 minutes.
3. The cages must be fully cooled prior to use.
4. Place the cages on the covered storage rack inside the animal room.